Xgal staining

This protocol assumes you already have fixed, hydrated and washed tissue. Heat fixation will not work for this protocol: use aldehyde fixation only. Tissues should not be treated with Methanol before Xgal staining, because Methanol inactivates the enzymatic activity of the β-Galactosidase enzyme.

1. Prepare Xgal staining buffer
2. Incubate tissue in staining buffer at 37°C (water bath or dry incubator are both fine) until you see blue staining. This may take anywhere from 15 minutes to overnight.

Xgal staining buffer: all you need to do is mix these things together – make up FRESH solution every time and take care of your stock solutions in the meantime

	original protocol volume	**original protocol stock concentrations**	**our stock concentrations (if different from original)**	**our protocol volumes**
	62 µl	50 mM potassium ferricyanide		62
	62 µl	50 mM potassium ferrocyanide		62
	15 µl	20% Triton X-100	10%	30
	30 µl	5M NaCl		30
	1 µl	1M MgCl2	50mM	20
	100 µl	10x PBS		100
	697 µl	water		675
	33 µl	8% Xgal in DMF (add this last)	50mg/ml	21
FINAL VOLUME	**1000 ul**			**1000 ul**