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*Proc. R. Soc. B* 2014 **281**, 20132673, published 5 February 2014

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## Research

**Cite this article:** Green II DA, Extavour CG. 2014 Insulin signalling underlies both plasticity and divergence of a reproductive trait in *Drosophila*. *Proc. R. Soc. B* **281**: 20132673. <http://dx.doi.org/10.1098/rspb.2013.2673>

Received: 14 October 2013

Accepted: 13 January 2014

### Subject Areas:

evolution, developmental biology, genetics

### Keywords:

ovariole, ovary, *Drosophila sechellia*, *Drosophila erecta*, phenotypic plasticity, insulin receptor

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Electronic supplementary material is available at <http://dx.doi.org/10.1098/rspb.2013.2673> or via <http://rspb.royalsocietypublishing.org>.

# Insulin signalling underlies both plasticity and divergence of a reproductive trait in *Drosophila*

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Phenotypic plasticity is the ability of a single genotype to yield distinct phenotypes in different environments. The molecular mechanisms linking phenotypic plasticity to the evolution of heritable diversification, however, are largely unknown. Here, we show that insulin/insulin-like growth factor signalling (IIS) underlies both phenotypic plasticity and evolutionary diversification of ovariole number, a quantitative reproductive trait, in *Drosophila*. IIS activity levels and sensitivity have diverged between species, leading to both species-specific ovariole number and species-specific nutritional plasticity in ovariole number. Plastic range of ovariole number correlates with ecological niche, suggesting that the degree of nutritional plasticity may be an adaptive trait. This demonstrates that a plastic response conserved across animals can underlie the evolution of morphological diversity, underscoring the potential pervasiveness of plasticity as an evolutionary mechanism.

## 1. Introduction

Phenotypic plasticity is the ability of a single genotype to yield distinct phenotypes in different environments. Phenotypic plasticity may play an important role in evolutionary diversification, as it is capable of generating striking examples of biodiversity, including differences in morphology, behaviour, life history and species interactions [1]. However, whether or not phenotypic plasticity promotes or impedes evolutionary diversification is still unclear, and has been under debate for decades [2]. One hypothesis is that common molecular mechanisms underlie both plasticity and interspecific variation in a trait, which would allow plasticity to promote diversification by providing a range of phenotypes whose underlying genetic variation can be subject to selection by genetic accommodation, genetic assimilation or other means [3]. The molecular underpinnings of plasticity within a single species are known for several systems [4–7], and there is also evidence that plasticity contributes to species differentiation [8]. However, specific examples that functionally demonstrate the molecular basis for both the plasticity and interspecies divergence of the same trait are lacking. We therefore sought to provide such an example, by examining the molecular basis of the evolutionary divergence and of the phenotypic plasticity of a single trait.

Reproductive traits are particularly relevant models for investigating the molecular mechanisms of phenotypic plasticity and evolutionary changes, because they affect the number of offspring, and hence fitness. Here, we examine one such trait: insect ovariole number. Ovarioles are egg-producing structures of insect ovaries. At the anterior end of each ovariole is the germarium, where germ-line stem cells (GSCs), supported within their somatic niche, self-renew and also differentiate to ultimately yield the mature oocyte and supporting germ cells. Posterior to the germarium, progressively maturing oocytes are arranged in an anterior to posterior progression within each ovariole.

Ovariole number spans three orders of magnitude across insects [9]. Several lines of evidence suggest that ovariole number is adaptive. First, ovariole number is a strong determinant of reproductive capacity, and thus is positively correlated to female fecundity and fitness [10–13]. Second, ovariole number is

heritable and lineage-specific. Quantitative and developmental genetic analyses suggest that inter- and intraspecies variations in ovariole number are controlled through multiple loci [14–21]. Third, ovariole number shows latitudinal and altitudinal clinal variation on multiple continents [22,23]. In two cosmopolitan *Drosophila* species, *D. melanogaster* and *D. simulans*, ovariole number is greater in temperate populations than in tropical populations [24]. Finally, ovariole number is correlated with species ecology. Low ovariole numbers commonly evolve among ecological specialists, whereas generalists, or insects with more heterogeneous food sources, tend to evolve higher ovariole numbers [25–28].

Ovariole number exhibits strong phenotypic plasticity in response to larval rearing environment, particularly nutrition [29,30] and temperature [31]. Previous attempts to relate genetically fixed variation in and phenotypic plasticity of ovariole number in *Drosophila* concluded that different developmental mechanisms were responsible for species-specific ovariole number and ovariole number plasticity [29]. However, the underlying molecular mechanisms remained unknown. Many developmental genetic details underlying ovariole number determination have since emerged [32–34], allowing for molecular investigations of the basis for ovariole number determination and divergence. In the following paragraph, we describe the essential cellular behaviours involved in ovariole morphogenesis. These developmental events suggest specific candidate processes and molecular mechanisms that may underlie the evolution of variation in ovariole number.

Ovary morphogenesis in *Drosophila* begins with the specification of somatic gonad precursor cells in late embryogenesis [35]. Unlike most larval tissues in *Drosophila*, somatic ovarian cells proliferate continuously throughout larval life with no dramatic cell death [36]. Ovariole morphogenesis begins with the stacking of somatic ovarian cells into structures called terminal filaments (TFs) in the anterior of the larval ovary [37]. TF number at the larval–pupal (LP) transition stage directly determines adult ovariole number [29], and the number and morphogenesis of TF cells at LP stage determines TF number [32]. Somatic ovarian cells are then specified as anterior versus posterior cells, and a constant percentage of the anterior cells become TF cells [33]. Insulin and ecdysone signalling regulates TF cell number through modulating somatic ovarian cell proliferation, differentiation and morphogenesis [33,38,39]. This suggests that variation in hormonal signalling could underlie one or both of species-specific ovariole number and the phenotypic plasticity of ovariole number.

Here, we examine the role of insulin/insulin-like growth factor signalling (IIS) in the determination of mean ovariole number and the phenotypic plasticity of ovariole number in *Drosophila*. Furthermore, we use a comparison of two *Drosophila* species, *D. melanogaster* and *D. sechellia*, to investigate the hypothesis that the same molecular mechanism regulates both species-specific values and phenotypic plasticity of the same trait, ovariole number.

## 2. Methods

### (a) *Drosophila* strains, culture conditions and diet manipulations

The following strains were used as wild-type strains for species comparisons: *D. melanogaster* OregonR-C (Bloomington *Drosophila*

Stock Center (BDSC) no. 5, gift of the Hartl laboratory, Harvard University); *D. sechellia* Robertson strain (UC San Diego *Drosophila* Species Stock Center (DSSC) no. 14021-0248.25, gift of the Hartl laboratory); *D. simulans* (DSSC no. 14021-0251.194); and *D. erecta* (DSSC no. 14021-0224.01). To evaluate the amount of intraspecies variation in ovariole number, we counted adult ovariole number in isofemale lines of *D. melanogaster* and *D. sechellia*. Both tropical and temperate populations of *D. melanogaster* were considered. Tropical *D. melanogaster* isofemale lines, established from a population in Zambia, were a gift of the Flatt Laboratory (University of Lausanne). North American *D. melanogaster* isofemale lines (derived from females collected in Nevada, Catalina Island, CA, Santa Fe, NM and Raleigh, NC) were a gift of the DePace laboratory (Harvard Medical School). *D. sechellia* isofemale lines were a gift of the Hartl laboratory.

To examine IIS function in *D. melanogaster*, the following lines were used: the *InR*<sup>339</sup> hypomorphic allele [40,41], a gift of the Hafen laboratory (ETH Zurich); the *InR*<sup>GC25</sup> inversion allele ([42], BDSC no. 9554); and the *Df(3R)Exel6186* deficiency allele (BDSC no. 7647).

To determine the role of systemic IIS from brain-derived peptides, we genetically ablated the principal insulin-producing cells of the brain. We used the *dilp2-Gal4* driver [43], which is expressed specifically in the paired small clusters of medial neurosecretory cells that are known to produce *Drosophila* insulin-like peptides. We crossed this driver to the *UAS-rpr* (BDSC no. 5824) line to drive expression of the proapoptotic gene *reaper*.

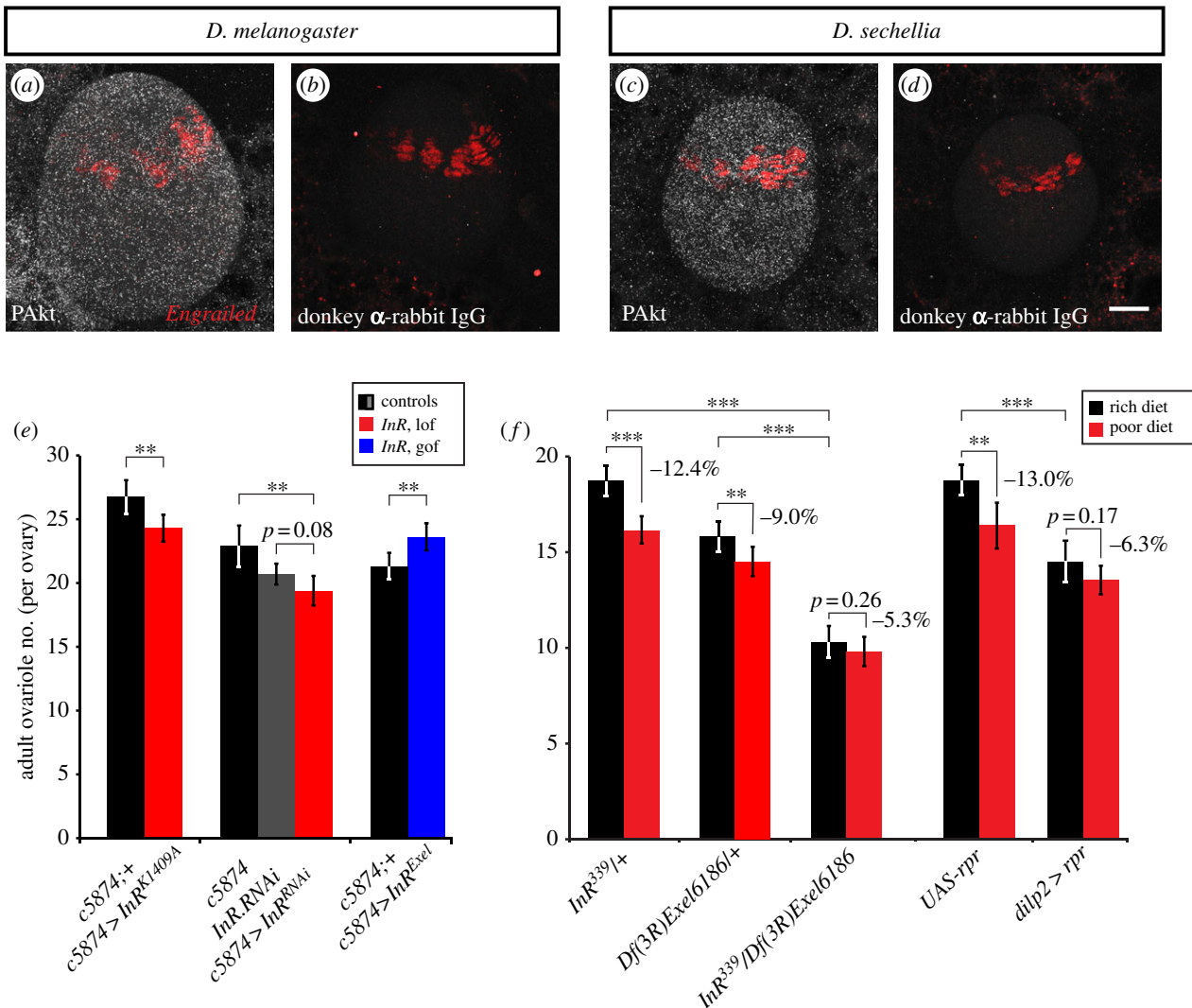
To determine the responsiveness of somatic ovarian cells to IIS, we altered expression of the *Drosophila* insulin-like receptor *InR* specifically in the ovary by using the *c587-GAL4* driver, which is expressed specifically in somatic ovarian cells beginning in the third larval instar ([44], gift of the Drummond-Barbosa laboratory, Johns Hopkins University). We crossed this driver to the following *UAS* lines to alter *InR* activity: *UAS-InR<sup>Exel</sup>* (BDSC no. 8262), *UAS-InR<sup>K1409A</sup>* (BDSC no. 8259) and *UAS-InR<sup>RNAi</sup>* (BDSC no. 31037).

All adult ovariole counts and LP transition stage TF counts were performed as previously described [32]. Tibia length (adult females) was used as an adult body size proxy, as it has been previously demonstrated to correlate positively with body mass, which is indicative of overall body size [45].

Flies were maintained on standard laboratory diet (32 g Torula yeast, 60.5 g corn meal, 128 g dextrose, 9.2 g agar per litre). In all diet manipulation experiments, flies were raised from egg through to adult on the specified diet. Rich diet for all analyses consisted of standard laboratory diet supplemented with active dry yeast. Poor diet consisted of standard laboratory diet diluted with 3% agar in a ratio of 1 : 3 (25% final concentration standard laboratory diet) with no dry yeast supplementation. Wortmannin (EMD Millipore) was dissolved in 100% methanol and added to standard laboratory diet at 1% v/v. All rearing and experiments were performed at 25°C and 60–70% humidity.

### (b) Quantitative PCR

As one measure of IIS pathway activity, we measured levels of *Thor* transcript [46]. Total RNA was extracted from ten biological replicates of five whole wandering third-instar females that were grown on rich diet. RNA was extracted using Trizol (Invitrogen), treated with TURBO DNase-I (Ambion, Life Technologies), and phenol–chloroform extracted. cDNA was prepared using oligo-dT primers and 0.5 µg RNA per reaction with Superscript III First Strand Synthesis Kit (Invitrogen). qPCR was performed using PerfeCta SYBR Green SuperMix, Low Rox (Quanta Biosciences). *gapdh1* was used to normalize RNA levels and *rp49* was used as expression control. Primer pairs were designed for use with both species templates. Primers were verified by performing species-specific standard curves for each primer pair, and showed less than 2.5% difference in amplification efficiency



**Figure 1.** Insulin signalling regulates ovariole number determination and plasticity in *Drosophila*. (a–d) IIS activity in larval ovaries of both *D. melanogaster* and *D. sechellia* visualized by phosphorylated Akt (pAkt; white; maximum projection of optical sections through whole ovary). *Engrailed* (red) marks terminal filament precursor cells. (b,d) Secondary antibody-only controls. Scale bar, 20  $\mu\text{m}$ . (e) Adult ovariole number in females with ovary-specific expression of *InR* alleles driven by the *c587*-GAL4 driver.  $n \geq 20$  ovaries for all genotypes. For *InR<sup>K1409A</sup>* and *InR<sup>Exel</sup>*, controls are siblings carrying a balancer chromosome (black bars). lof, loss of function; gof, gain of function. (f) Adult ovariole number in females with loss of *InR* function (*InR<sup>339</sup>/Df(3R)Exel6186*) or with *dilP*-producing neurons ablated (*dilp2-GAL4>UAS:rpr*), reared on rich or poor diets.  $n \geq 20$  for all genotypes. In (e,f), error bars show 95% CI of means. Student's *t*-test: \*\*\* $p < 0.001$ , \*\* $p < 0.01$ .

between species. Primer pairs were as follows: *gapdh1-f*, AGCC-GAGTATGTGGTGGAGT, *gapdh1-r*, GGCTGTAGGCGTCCAGGTTA; *Thor-f*, AGCTAAGATGTCGGCTTCACC, *Thor-r*, TTTGGTGCCCTCCAGGAGTGG; *rp49-f*, TGCTAAGCTGTCCGACAAATG, *rp49-r*, TICTTGAATCCGGTGGGCAG.

### (c) Immunohistochemistry, confocal imaging and analysis

Immunostaining was carried out as previously described [32]. The following primary antibodies were used: mouse 4D9 anti-*Engrailed* (1:40, Developmental Studies Hybridoma Bank); rabbit anti-Vasa (1:500, gift of P. Lasko, McGill University); and rabbit anti-phospho-*Drosophila* Akt (Ser505) (1:200, Cell Signaling Technology no. 4054). Secondary reagents used were Hoechst 33342 (Sigma, 1:1000 of 10 mg ml<sup>-1</sup> stock solution), goat anti-mouse Alexa 488, goat anti-guinea pig Alexa 488 and donkey anti-rabbit Alexa 555 (1:500, Invitrogen). Samples were mounted in Vectashield (Vector laboratories) and imaged using a Zeiss LSM 780 confocal microscope.

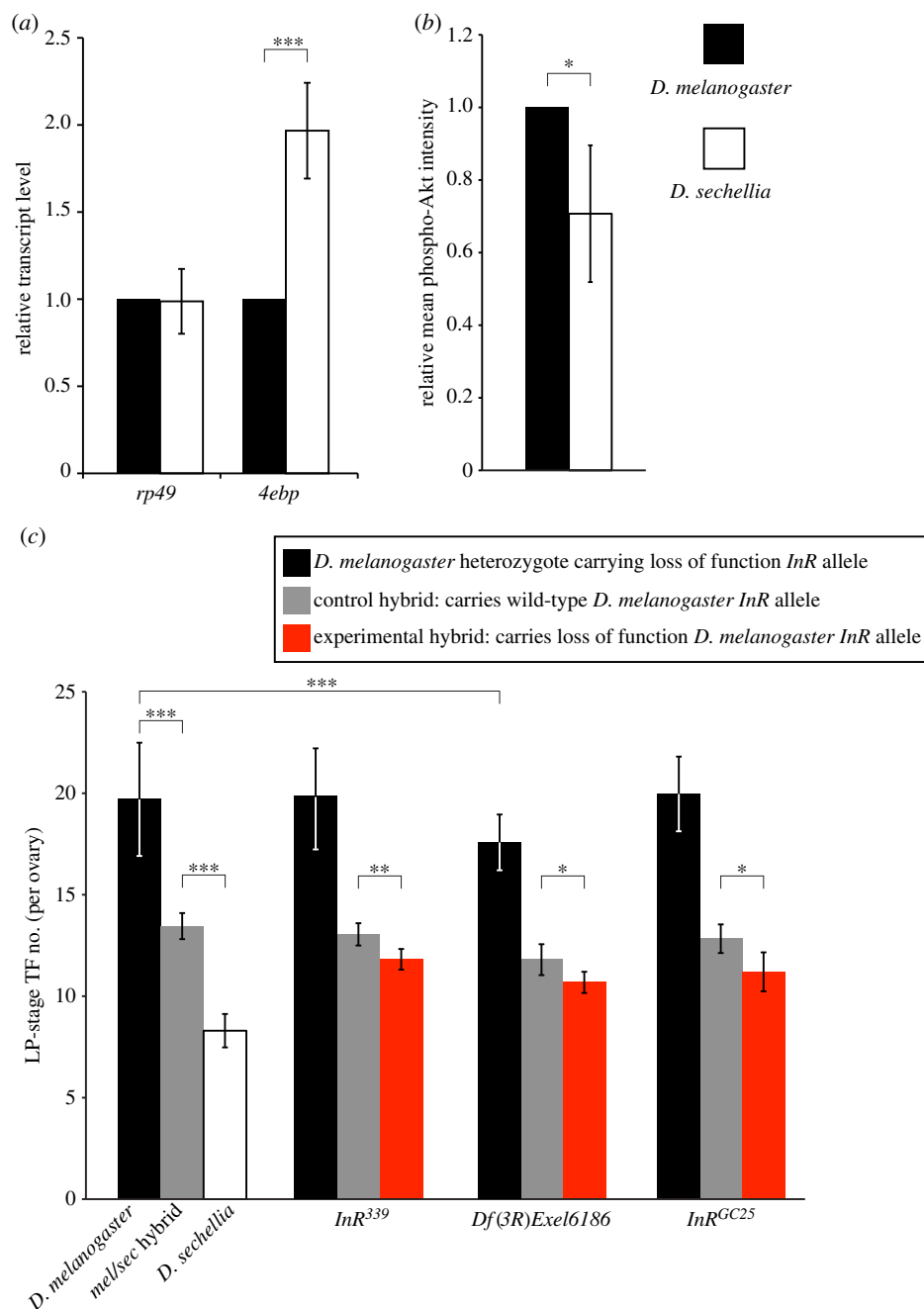
Phosphorylated Akt (phospho-Akt) staining was quantified by measuring mean fluorescence signal intensity from maximum projection images composed of an equal number of confocal z-slices

for each ovary. Secondary-only controls (figure 1b,d) indicate that the staining detected (figure 1a,c) and measured (figure 2b) is not background signal. A standard area of specifically anterior somatic ovarian cells, the cells from which TF precursor cells are specified, was analysed. Phospho-Akt intensity was normalized to mean DNA (Hoechst 33342) staining intensity to control for potential differences due to specific immunostaining experiments. Images were analysed with IMAGEJ v. 1.45I.

### (d) Statistical analyses

Student's *t*-test was used for all pairwise comparisons of differences in means unless otherwise noted. Bonferroni adjustment for multiple comparisons was performed as appropriate. Mann–Whitney (Wilcoxon) test was used to evaluate differences in phospho-Akt staining intensity. To evaluate differences in interpopulational variation in ovariole number, Bartlett's test was used, as this test does not assume homogeneity of the variance of species-specific variances. Homo-/heterogeneity of species-specific variances was tested with Welch ANOVA (Welch *t*) to account for differences in mean values. Correlations, where noted, were evaluated by least-squares linear regression of mean values for each genotype. Statistical analyses were performed in EXCEL and JMP PRO v. 11.





**Figure 2.** Differential IIS activity exists between *D. melanogaster* and *D. sechellia*. Quantified levels of (a) larval expression of the growth attenuator *Thor* (normalized to *gapdh1*) and the ribosomal protein gene *rp49* and (b) phospho-Akt intensity in wandering third-instar larval ovaries of *D. melanogaster* compared with *D. sechellia*. (c) Left-most set of bars shows TF number in *D. melanogaster* (black bar), *D. melanogaster*/*D. sechellia* hybrid (grey bar) and *D. sechellia* (white bar) females. Remaining sets show adult ovariole number in *D. melanogaster* females heterozygous for *InR* loss of function mutation (black bars), and final TF number in *D. melanogaster* *InR*<sup>+/−</sup>/*D. sechellia* hybrids (where *InR*<sup>\*</sup> is any of three different *InR* loss of function alleles; red bars) compared with *D. melanogaster* *InR*<sup>+/+</sup>/*D. sechellia* control hybrids (grey bars). Controls are sisters carrying a wild-type copy of *D. melanogaster* *InR*.  $n \geq 10$  for all genotypes. Error bars show 95% CI of means. Student's *t*-test in (a,c); Mann–Whitney (Wilcoxon) test in (b): \*\*\* $p < 0.001$ , \*\* $p < 0.005$ , \* $p < 0.05$ .

### 3. Results

#### (a) Role of systemic IIS in determining ovariole number

We previously showed that loss of function of the *Drosophila* insulin-like receptor (*InR*) in *D. melanogaster* significantly reduces TF number by reducing both the number of somatic gonad precursor cells and the subsequent somatic cell proliferation rate throughout larval life [33]. To determine whether TF number reduction in *InR* mutants is due to autonomous IIS activity in somatic ovarian cells rather than through an indirect mechanism, we first asked whether IIS is active in ovarian cells at the relevant developmental time.

Phosphorylated Akt (phospho-Akt) protein, an indicator of active IIS, was detectable at levels above background in wandering third larval instar ovaries, the time at which TF cells are proliferating and TFs are forming (figure 1*a–d*; compare *a* with *b*, and *c* with *d*). Phospho-Akt was also detected at above-background levels in the fat body, however at lower levels than in the ovary (figure 1*a–d*). We then used the somatic ovary-specific driver *c587-GAL4* to abrogate or increase IIS specifically in the ovary. When IIS was decreased in the ovary either with the dominant negative *InR* allele *K1409A*, or with an *InR* RNAi construct, ovariole number was significantly decreased ( $p < 0.01$  for *InR*<sup>K1409A</sup>,  $p < 0.01$

and  $p = 0.08$  for *c587-GAL4* and *UAS:InR<sup>RNAi</sup>* parental controls, respectively; figure 1e). Conversely, when IIS was increased with overexpression of wild-type *InR* (*InR<sup>Exel</sup>*), ovariole number was significantly increased ( $p < 0.01$ ; figure 1e). As expected owing to the use of an ovary-specific *GAL4* driver, these changes in ovariole number were not simple consequences of changes in body size (see electronic supplementary material, appendix S1). Finally, to determine whether systemic IIS from brain-derived insulin-like peptides (dILPs) regulates ovariole number determination, we genetically ablated insulin-producing neurons by using a *dilp2-GAL4* driver to overexpress the proapoptotic gene *reaper* (*rpr*). Adult ovariole number was significantly reduced in *dilp2 > rpr* females compared with *UAS:rpr* control females (figure 1f;  $p < 0.001$ ). Taken together, these results show that systemic IIS from brain-derived dILPs controls autonomous somatic ovarian cell proliferation, and modulation of IIS leads to changes in ovariole number.

### (b) Role of IIS in nutritional plasticity of ovariole number

Systemic IIS is nutritionally controlled [47]. To test whether IIS mediates the nutritional plasticity of ovariole number, we reared flies with wild-type or modulated levels of IIS on rich or poor diets (see Methods). Like in wild-type flies [32], ovariole numbers were significantly reduced by poor diet in heterozygotes for *InR* loss of function mutations or *UAS:rpr* controls (figure 1f;  $p < 0.001$  and  $p < 0.01$ , respectively). Body size of flies with altered IIS levels showed a more variable response to poor diet than wild-type flies, and body size was not a reliable predictor of ovariole number across genotypes (see electronic supplementary material, appendix S2). However, *InR* loss of function mutant and *dilp2 > rpr* females showed no statistically significant change in ovariole number on rich versus poor diet (figure 1f;  $p = 0.39$  and  $p = 0.17$ , respectively). These results show that IIS is a molecular mediator of nutritional plasticity of ovariole number in *D. melanogaster*.

### (c) IIS activity and sensitivity in *D. melanogaster* and *D. sechellia*

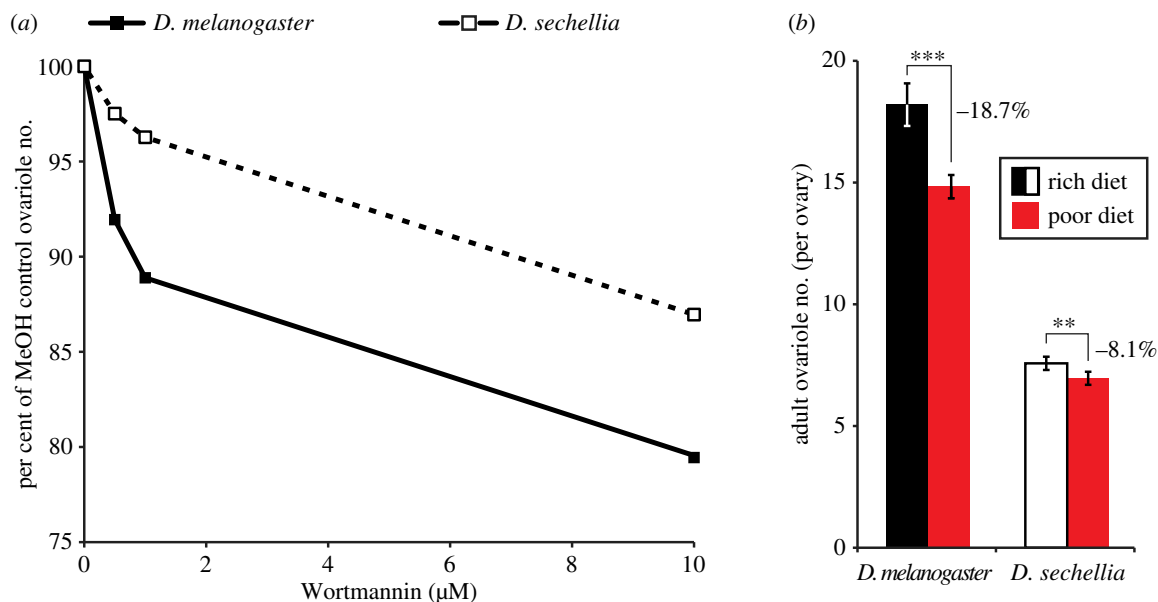
The *melanogaster* subgroup species *D. melanogaster* and *D. sechellia*, which diverged only five million years ago, have remarkably divergent mean ovariole numbers of 18.2 and 7.6, respectively. We previously showed that the heritable ovariole number difference between these species is caused by differences in somatic gonad precursor cell specification and somatic ovarian cell proliferation rate throughout larval life, and that *InR* loss of function mutants in *D. melanogaster* phenocopy both these differences [33]. We therefore hypothesized that IIS activity is reduced in *D. sechellia* compared with *D. melanogaster*. To test this hypothesis, we measured transcript expression of the growth attenuator *Thor*, which is negatively regulated by IIS [46]. 4E-BP, the protein product of the *Thor* transcript, is a known negative regulator of cell number in *Drosophila* [46]. We found that *Thor* expression in *D. sechellia* was significantly greater than in *D. melanogaster* (figure 2a;  $p < 0.001$ ). In addition, we quantified the levels of phospho-Akt in the larval ovary of both species, and found that these levels were significantly higher in ovaries of *D. melanogaster* than of *D. sechellia* (figure 2b;  $p < 0.05$ ). Taken together, these assessments of IIS activity indicate that IIS operates at higher

levels in *D. melanogaster* than in *D. sechellia*. Consistent with these results, body size of *D. sechellia* is significantly smaller than that of *D. melanogaster* (see electronic supplementary material, appendix S4;  $p < 0.001$ ). This suggests that evolutionary changes in IIS contribute to the divergence in ovariole number between these two species.

To further test for species-specific differences in IIS-mediated control of TF number between *D. melanogaster* and *D. sechellia*, we used interspecies hybrid complementation tests. Previous quantitative genetics analysis suggested that the *InR* locus may contribute to interspecies variation in ovariole number [15]. Therefore, we crossed *D. melanogaster* females carrying *InR* loss of function mutations with *D. sechellia* males, and counted TF number in resulting *melanogaster/sechellia* hybrids. Hybrids carrying mutant *InR* alleles from *D. melanogaster* had significantly reduced body size (see electronic supplementary material, appendix S3) and TF number compared with control hybrids carrying a wild-type *D. melanogaster InR* allele (figure 2c;  $p < 0.05$  for *InR<sup>GC25</sup>* and *Df(3R)Exel6186*;  $p < 0.01$  for *InR<sup>339</sup>*). This suggests that the wild-type *D. melanogaster InR* allele may confer a higher level of IIS than the *D. sechellia* allele.

Because *D. sechellia InR* mutants are not available, we could not test this hypothesis directly by creating hybrids carrying a loss of function *D. sechellia InR* allele and a wild-type *D. melanogaster InR* allele. However, if our interpretation is correct, then the decrease in ovariole number caused by loss of one functional *D. melanogaster InR* allele should be less severe in *D. melanogaster* heterozygotes than in *melanogaster/sechellia* hybrids. Consistent with our hypothesis, adult ovariole number in *D. melanogaster InR* loss of function heterozygotes was not significantly different from wild-type (*Oregon R*) for two different *InR* alleles, *InR<sup>GC25</sup>* and *InR<sup>339</sup>* (figure 2c;  $p = 0.20, 0.34$ , respectively). For a third *D. melanogaster InR* loss of function allele, *Df(3R)Exel6186*, adult ovariole number was significantly lower than wild-type (figure 2c;  $p < 0.001$ ), but the degree of reduction in ovariole was somewhat lower than that seen in the *melanogaster/sechellia* hybrid for the same *InR* allele (12.5% versus 13.6% reduction in ovariole number; figures 1f and 2c). In summary, with these experiments, we have compared the decrease in ovariole number caused by heterozygosity for a loss of function *D. melanogaster InR* allele in *D. melanogaster* heterozygotes versus *D. melanogaster/D. sechellia* hybrids, and shown that the ovariole number decrease is higher in the interspecies hybrids (figure 2c). Overall, these results are consistent with our hypothesis that the wild-type *D. melanogaster InR* allele confers a higher level of IIS than the wild-type *D. sechellia* allele, consistent with IIS activity being higher in *D. melanogaster* compared with *D. sechellia*. Taken together, these data demonstrate that IIS activity differs between *D. melanogaster* and *D. sechellia*, and that this activity difference contributes to species-specific ovariole number.

IIS sensitivity controls differential plastic response to nutrition in several insect species [48–50]. To determine how IIS activity difference could influence nutrition-dependent plasticity of ovariole number, we fed flies food containing Wortmannin, a specific inhibitor of PI3K [51], in a graded concentration series. Consistent with the results of genetic manipulation of IIS (figures 1e,f and 2c), body size (see electronic supplementary material, appendix S4) and ovariole number (figure 3a) were reduced in a dose-dependent manner in both species when grown on food containing Wortmannin. However, at all tested concentrations of Wortmannin, ovariole number was more significantly reduced in *D. melanogaster*



**Figure 3.** Differential IIS sensitivity between *D. melanogaster* and *D. sechellia* correlates with species-specific nutritional plasticity of ovariole number. (a) Dose-dependent Wortmannin-induced decrease in adult ovariole number in *D. melanogaster* and *D. sechellia*, shown as percentage decrease relative to flies reared on control food containing methanol (Wortmannin solvent).  $n = 20$  for each species at all concentrations except *D. sechellia* at  $10 \mu\text{M}$ ,  $n = 6$  (owing to low eclosion rate of *D. sechellia* at high Wortmannin concentrations). (b) Adult ovariole number in *D. melanogaster* and *D. sechellia* reared on poor and rich diets. Error bars show 95% CI of means.  $n \geq 20$  for all genotypes and conditions. Student's  $t$ -test: \*\*\* $p < 0.001$ , \*\* $p < 0.005$ . (Online version in colour.)

than in *D. sechellia* (figure 3a). This indicates that ovariole number is more sensitive to changes in IIS in *D. melanogaster* than in *D. sechellia*. Together with our finding of evolved differences in IIS between the two species, this also shows that higher IIS activity in *D. melanogaster* is correlated with higher sensitivity to changes in IIS compared with *D. sechellia*.

#### (d) Correlation between IIS sensitivity and nutritional plasticity

To test whether evolved differences in IIS activity levels and sensitivity could yield differences in nutritional plasticity between species, we measured ovariole number nutritional plasticity for *D. sechellia*. As in *D. melanogaster* [32], poor diet reduced ovariole number in *D. sechellia*, but only by 8.1%, in contrast to 18.7% in *D. melanogaster* (figure 3b). Body size was significantly reduced by poor diet in *D. melanogaster* ( $p < 0.001$ ), whereas in *D. sechellia* body size was reduced numerically but not significantly ( $p = 0.08$ ; electronic supplementary material, appendix S5). These data demonstrate that evolutionary change in IIS underlies the divergence of both mean ovariole number and the nutritional plasticity of ovariole number between these two *Drosophila* species.

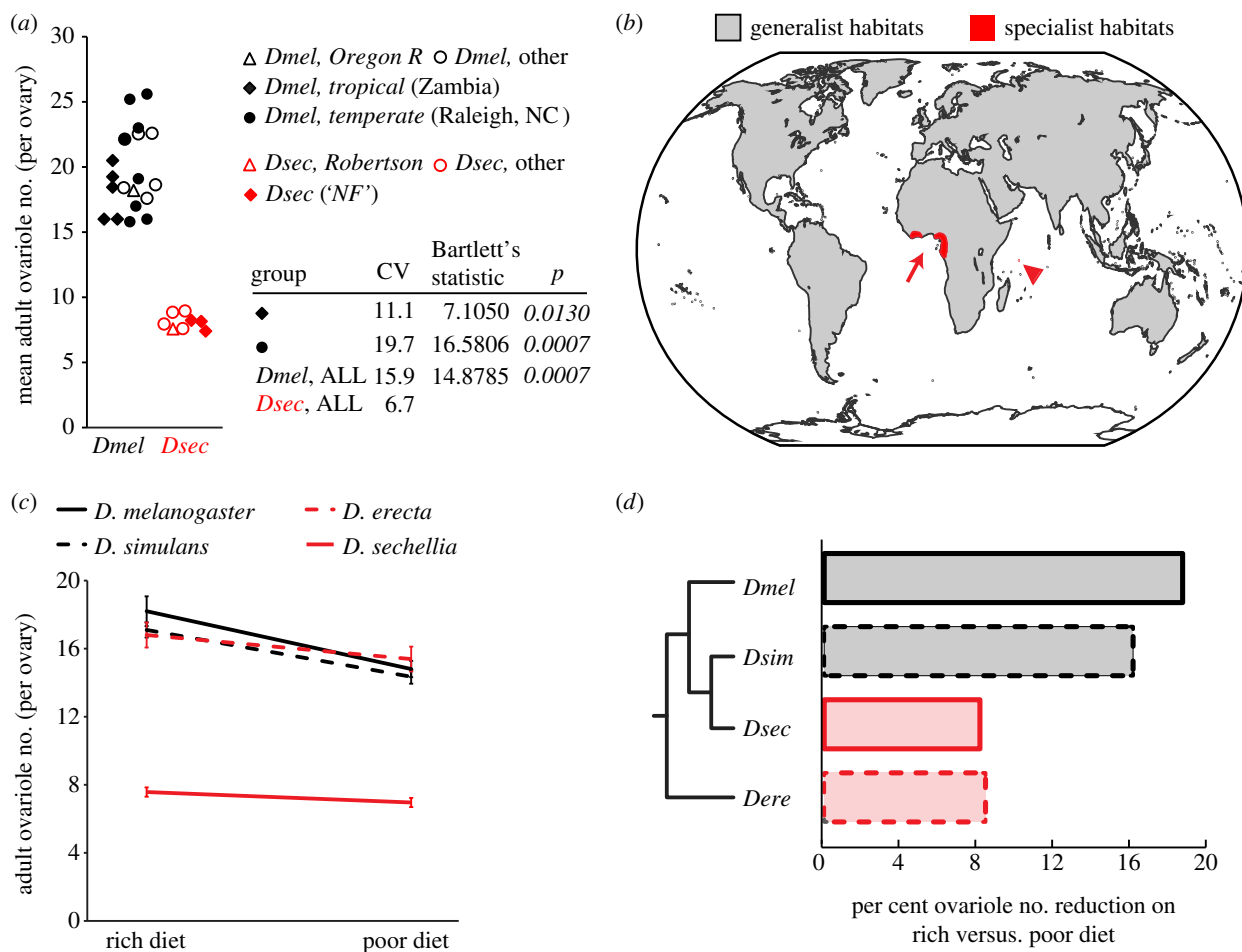
#### (e) Interpopulational variation in ovariole number

If plasticity promotes diversification by providing a range of phenotypes whose underlying genetic variation can be subject to selection, then modulating the degree of plasticity may lead to differences in interpopulational divergence. Having observed that the degree of nutritional plasticity in ovariole number has diverged between *D. melanogaster* and *D. sechellia*, we asked whether interpopulational variation in ovariole number also differs between these species. We measured mean ovariole number for multiple isofemale lines from both species, and observed greater between-population variation for ovariole number in *D. melanogaster*

compared with *D. sechellia* (figure 4a and the electronic supplementary material, appendix S6;  $p < 0.001$ ). Although *D. sechellia* occupies an exclusively tropical habitat whereas *D. melanogaster* is distributed worldwide (figure 4b), even when considering variation within a tropical *D. melanogaster* population, variation is significantly greater in *D. melanogaster* compared with *D. sechellia* (figure 4a,  $p < 0.05$ ). Genetic variation in *D. sechellia* is known to be lower than that of other *melanogaster* group species [52], and it is possible that this contributes to its reduced interpopulational variation in ovariole number. However, we argue that IIS-dependent plasticity provides a proximate molecular mechanism for the evolutionary divergence of ovariole number. Our data are consistent with the idea that plasticity plays a central role in diversifying ovariole number not only between species, but also within species.

#### (f) Correlation between ecology and nutritional plasticity of ovariole number

Finally, we asked whether nutrition-dependent plasticity of this critical reproductive trait was linked to broader ecological patterns of ovariole number diversity, which may indicate an adaptive value of nutritional plasticity. Mean ovariole number is correlated with nutritional host preference in many insect species from a range of global habitats [25–27]. Specifically, species that have a wide host preference (generalists) or feed on abundant food sources tend to have more ovarioles than species that feed on a restricted niche (specialists) or scarce food sources. This correlation has been used to support the idea of adaptive value of ovariole number in terms of  $r$ - $K$  selection theory [11,25,28]. Briefly, higher ovariole numbers permitting a larger number of offspring ( $r$ -selection) would be favoured when host substrates are numerous and nutritionally rich, whereas restricted substrates would favour production of fewer offspring and hence decreased ovariole number ( $K$ -selection). Given our finding that mean ovariole number and nutritional plasticity of ovariole number are controlled by the



**Figure 4.** Degree of plasticity correlates with relative interpopulational variation and ecological niche. (a) Range of ovariole number in different populations of *D. melanogaster* and *D. sechellia*. CV = coefficient of variation. Bartlett's test was used to compare variance of all *D. sechellia* populations with variance of indicated *D. melanogaster* groups. *p*-Values indicating a significant difference are italicized. (b) Global species distributions of the generalists *D. melanogaster* and *D. simulans* (worldwide distributions: grey); the specialists *D. erecta* (red arrow: specializes on fruits of the genus *Pandanus* in west Africa) and *D. sechellia* (red arrowhead: specializes on the fruit *M. citrifolia* in the Seychelles, which is toxic to other *Drosophila* species) have limited habitats. (c) Reaction norm of ovariole number on rich versus poor diet in four *Drosophila* species. Poor diet reduces ovariole number more in cosmopolitan generalist species (black lines) than in specialist species (red lines). Error bars show 95% CI of means.  $n \geq 20$  for all genotypes and conditions. (d) The phylogenetic relationship between the four species analysed is shown to the left of a bar graph showing per cent ovariole number reduction on different diets.

same molecular mechanisms, we predicted that nutritional plasticity would also correlate with range of host preference. Consistent with our hypothesis, we found that the cosmopolitan generalist species *D. melanogaster* and *D. simulans* (figure 4b) show high ovariole number plasticity and moderate body size plasticity in response to nutrition, whereas the specialist species *D. sechellia* [53] and *D. erecta* [54] show low nutritional plasticity and no significant change in body size (figure 4c,d and the electronic supplementary material, appendix S5). Because all species were reared on standard laboratory medium rather than native diets, we cannot rule out the possibility that our observed ovariole numbers and associated phenotypes may be affected by the use of a standard, non-native diet that was necessary to allow us to make comparisons between species. We note, however, that in the case of *D. sechellia*, ovariole number reported here is the same as that reported when *D. sechellia* is reared on its host plant *Morinda citrifolia* [53], suggesting that it may indeed be the degree of food source specialization, rather than a specific food source, which is the relevant parameter influencing ovariole number and its plasticity. Furthermore, low nutritional plasticity in *D. erecta*, which specializes on the non-toxic *Pandanus* genus of plants, indicates that this effect is not an artefact of the toxicity of *M. citrifolia* to

other *Drosophila* species. Our experiments thus demonstrate that *Drosophila* species differ in their sensitivity to nutritional input, and suggest that relative IIS activity level may mediate these sensitivity differences.

Differences in plasticity lead to different relative ovariole numbers, and hence different relative reproductive capacities, between species in different environments (figure 4c). These results imply that the degree of nutritional plasticity in ovariole number may be subject to selection, and has diverged across species in response to ecological niche. If specific nutritional plasticity is an adaptation to host preference range, then variation in IIS levels and sensitivity could provide a proximate mechanism for the observed correlation between mean ovariole number and host preference.

## 4. Discussion

Ovariole number is believed to be under stabilizing selection [17], and environmental changes cannot increase ovariole number beyond a lineage-specific maximum [10,55]. Evolution of reduced ovariole number has occurred convergently in many insect lineages [33], and is correlated with occupation of



specialist ecological and nutritional niches [25–27]. Consistent with these observations, we suggest that nutritional plasticity and reproductive capacity may present a trade-off dependent on relative IIS activity: high IIS activity can increase mean ovariole number, but at the cost of strongly reducing ovariole number in poor nutritional conditions (figure 4c). Because increased IIS also correlates with shortened lifespan [56,57], it is also possible that evolution of low plasticity due to low IIS levels could confer the advantage of an increased lifespan that is relatively robust to changes in nutritional conditions. Although we cannot yet determine which of these traits is the target of selection, we suggest that evolutionary diversification of both ovariole number and its nutritional plasticity occurs through genetic changes that modulate IIS activity and sensitivity. Our data show that a functional consequence of evolutionary changes in IIS activity and sensitivity is modulation of plastic range between species, and that this range is correlated with interpopulational diversification. We previously showed that different developmental mechanisms, which are genetically separable, contribute to ovariole number evolution [33]. We hypothesize that these alternative mechanisms may be targets of evolution for generating population-specific ovariole number while maintaining species-specific plastic responses.

While we have demonstrated that IIS has diverged between *Drosophila* species, what remains to be elucidated are the specific loci responsible for this divergence. Our data, particularly the interspecies hybrid complementation results, are consistent with the hypothesis that evolutionary change at the *InR* locus contributes to interspecies variation in ovariole number. Cross-species transgenesis and in-depth genetic analysis of IIS differences between species will be necessary to address this problem. We note here that both coding and noncoding differences exist between *D. melanogaster* and *D. sechellia* at the *InR* locus, none of which suggest obvious candidates for functional divergence. The protein coding sequences are 97% identical between these two species, and none of the amino acid changes occur within the known kinase domain. This suggests that slight structural or non-

kinase activity-related alterations in the *InR* protein could modulate signalling in such a way as to contribute to phenotypic change. Natural variation in a coding region indel polymorphism in *InR* among *D. melanogaster* populations is consistent with this hypothesis [58].

IIS in multicellular animals is a conserved mechanism that coordinates cellular growth and proliferation with physiological condition, particularly nutritional state. The regulation of IIS contributes to evolutionary change within invertebrate and vertebrate species [49,59]. We have now shown that the regulation of IIS can underlie evolutionary morphological diversity both within and between species. Interestingly, evidence from functional studies in *D. melanogaster* and in horned beetles suggests that both increasing [48] and decreasing [49] IIS can reduce nutritional plasticity. This suggests that IIS may be able to act as a nutritional stress response system that is either environment-sensitive or environment-insensitive. Ovariole number in *Drosophila* (this study) and ornament size in horned beetles [49] appear to be examples of environment-sensitive nutritional stress responses, allowing generation of more offspring or exaggerated ornaments when food is plentiful, and restricting investment in these traits when food is scarce. An example of environment-insensitive nutrient stress response may be external genitalia in *Drosophila*, which continue to devote resources to growth despite unfavourable environmental conditions [48]. Given the wide conservation of IIS-mediated growth response, this work suggests a potentially pervasive role of plasticity in generating adaptive diversity.

**Acknowledgements.** D.A.G. is supported by an NSF pre-doctoral fellowship and a Ford Foundation Dissertation Fellowship. This work was partially supported by NIH grant 1R01-HD073499. The authors declare no competing financial interests. D.A.G. and C.G.E. designed research; D.A.G. performed experiments and collected data; D.A.G. and C.G.E. analysed data and wrote the paper; C.E. obtained funding for the research. We thank Julien Ayroles, Ernst Hafen, Daniel Hartl, Paul Lasko, Peter Klepsatel, Thomas Flatt, Akhila Rajan, Norbert Perrimon, Sarah Saminadin-Peter and Angela DePace for reagents, the Extavour laboratory for discussion, and Yuichiro Suzuki and James Mallet for comments on the manuscript.

## References

- Moczek AP, Sultan S, Foster S, Ledon-Rettig C, Dworkin I, Nijhout HF, Abouheif E, Pfennig DW. 2011 The role of developmental plasticity in evolutionary innovation. *Proc. R. Soc. B* **278**, 2705–2713. (doi:10.1098/rspb.2011.0971)
- West-Eberhard MJ. 2003 *Developmental plasticity and evolution*, p. 794. Oxford, UK: Oxford University Press.
- Waddington CH. 1942 Canalization of development and the inheritance of acquired characters. *Nature* **3811**, 563–565. (doi:10.1038/150563a0)
- Brakefield PM, Gates J, Keys D, Kesbeke F, Wijngaarden PJ, Monteiro A, French V, Carroll SB. 1996 Development, plasticity and evolution of butterfly eyespot patterns. *Nature* **384**, 236–242. (doi:10.1038/384236a0)
- Rutherford AL, Lindquist S. 1998 Hsp90 as a capacitor for morphological evolution. *Nature* **396**, 336–342. (doi:10.1038/24550)
- Suzuki Y, Nijhout HF. 2006 Evolution of a polyphenism by genetic accommodation. *Science (New York, NY)* **311**, 650–652. (doi:10.1126/science.1118888)
- Abouheif E, Wray GA. 2002 Evolution of the gene network underlying wing polyphenism in ants. *Science* **297**, 249–252. (doi:10.1126/science.1071468)
- Bloom S, Ledon-Rettig C, Infante C, Everly A, Hanken J, Nascone-Yoder N. 2013 Developmental origins of a novel gut morphology in frogs. *Evol. Dev.* **15**, 213–223. (doi:10.1111/ede.12035)
- Bünning J. 1994 *The insect ovary: ultrastructure, previtellogenic growth and evolution*, p. 400. London, UK: Chapman and Hall.
- Cohet Y, David JR. 1978 Control of the adult reproductive potential by preimaginal thermal conditions. *Oecologia* **36**, 295–306. (doi:10.1007/BF00348055)
- Bouletreau-Merle J, Allemand R, Cohet Y, David JR. 1982 Reproductive strategy in *Drosophila melanogaster*: significance of a genetic divergence between temperate and tropical populations. *Oecologia* **53**, 323–329. (doi:10.1007/BF00389008)
- David JR. 1970 Le nombre d'ovarioles chez *Drosophila melanogaster*: relation avec la fécondité et valeur adaptive. *Arch. Zool. Exp. Gen.* **111**, 357–370.
- Klepsatel P, Galikova M, De Maio N, Ricci S, Schlotterer C, Flatt T. 2013 Reproductive and post-reproductive life history of wild-caught *Drosophila melanogaster* under laboratory conditions. *J. Evol. Biol.* **26**, 1508–1520. (doi:10.1111/jeb.12155)
- Bergland AO, Genissel A, Nuzhdin SV, Tatar M. 2008 Quantitative trait loci affecting phenotypic plasticity and the allometric relationship of ovariole number and thorax length in *Drosophila melanogaster*. *Genetics* **180**, 567–582. (doi:10.1534/genetics.108.088906)
- Orgogozo V, Broman KW, Stern DL. 2006 High-resolution quantitative trait locus mapping reveals sign epistasis controlling ovariole number between two *Drosophila* species. *Genetics* **173**, 197–205. (doi:10.1534/genetics.105.054098)
- Wayne ML, Hackett JB, Mackay TFC. 1997 Quantitative genetics of ovariole number in *Drosophila melanogaster*.

- I. Segregating variation and fitness. *Evolution* **4**, 1156–1163. (doi:10.2307/2411045)
17. Wayne ML, Mackay TFC. 1998 Quantitative genetics of ovariole number in *Drosophila melanogaster*. II. Mutational variation and genotype–environment interaction. *Genetics* **148**, 201–210.
18. Wayne ML, Hackett JB, Dilda CL, Nuzhdin SV, Pasyukova EG, Mackay TF. 2001 Quantitative trait locus mapping of fitness-related traits in *Drosophila melanogaster*. *Genet. Res.* **77**, 107–116. (doi:10.1017/S0016672300004894)
19. Wayne ML, McIntyre LM. 2002 Combining mapping and arraying: an approach to candidate gene identification. *Proc. Natl Acad. Sci. USA* **99**, 14 903–14 906. (doi:10.1073/pnas.222549199)
20. Telonis-Scott M, Bono L, McIntyre L, Wayne M. 2005 Analyses of the quantitative genetic architecture of ovariole number and body size in *Drosophila melanogaster* using diallels. In *46th Annual Drosophila Conf., San Diego, CA, 30 March–3 April 2005*, p. 806B.
21. Telonis-Scott M, McIntyre LM, Wayne ML. 2005 Genetic architecture of two fitness-related traits in *Drosophila melanogaster*: ovariole number and thorax length. *Genetica* **125**, 211–222. (doi:10.1007/s10709-005-8549-4)
22. Collinge JE, Hoffmann AA, McKechnie SW. 2006 Altitudinal patterns for latitudinally varying traits and polymorphic markers in *Drosophila melanogaster* from eastern Australia. *J. Evol. Biol.* **19**, 473–482. (doi:10.1111/j.1420-9101.2005.01016.x)
23. Wayne ML, Korol A, Mackay TF. 2005 Microclinal variation for ovariole number and body size in *Drosophila melanogaster* in 'evolution canyon'. *Genetica* **123**, 263–270. (doi:10.1007/s10709-004-5056-y)
24. Capy P, Pla E, David JR. 1993 Phenotypic and genetic variability of morphometrical traits in natural populations of *Drosophila melanogaster* and *D. simulans*. I. Geographic variations. *Genet. Sel. Evol.* **25**, 517–536. (doi:10.1186/1297-9686-25-6-517)
25. Kambyzellis MP, Heed WB. 1971 Studies of oogenesis in natural populations of Drosophilidae. I. Relation of ovarian development and ecological habitats of the Hawaiian species. *Am. Nat.* **94**(1), 31–49. (doi:10.1086/282700)
26. Fitt G. 1990 Variation in ovariole number and egg size of species of *Dacus* (Diptera: Tephritidae) and their relation to host specialization. *Ecol. Entomol.* **15**, 255–264. (doi:10.1111/j.1365-2311.1990.tb00807.x)
27. Leather S, Wellings P, Walters K. 1988 Variation in ovariole number within the Aphidoidea. *J. Nat. Hist.* **22**, 381–393. (doi:10.1080/00222938800770271)
28. Montague JT, Mangan RL, Starmer WT. 1981 Reproductive allocation in the Hawaiian Drosophilidae: egg size and number. *Am. Nat.* **118**, 865–871. (doi:10.1086/283877)
29. Hodin J, Riddiford LM. 2000 Different mechanisms underlie phenotypic plasticity and interspecific variation for a reproductive character in Drosophilids (Insecta: Diptera). *Evolution* **5**, 1638–1653.
30. Tu MP, Tatar M. 2003 Juvenile diet restriction and the aging and reproduction of adult *Drosophila melanogaster*. *Aging Cell* **2**, 327–333. (doi:10.1046/j.1474-9728.2003.00064.x)
31. Delpuech J-M, Noreteau B, Chiche J, Pla E, Vouldibio J, David JR. 1995 Phenotypic plasticity and reaction norms in temperate and tropical populations of *Drosophila melanogaster*: ovarian size and developmental temperature. *Evolution* **4**, 670–675. (doi:10.2307/2410320)
32. Sarikaya D, Assefa Belay A, Ahuja A, Green AD, Dorta A, Extavour CG. 2012 The roles of cell size and cell number in determining ovariole number in *Drosophila*. *Dev. Biol.* **363**, 279–289. (doi:10.1016/j.ydbio.2011.12.017)
33. Green II DA, Extavour CG. 2012 Convergent evolution of a reproductive trait through distinct developmental mechanisms in *Drosophila*. *Dev. Biol.* **372**, 120–130. (doi:10.1016/j.ydbio.2012.09.014)
34. Bartoletti M *et al.* 2012 Genetic basis for developmental homeostasis of germline stem cell niche number: a network of Tramtrack-Group nuclear BTB factors. *PLoS ONE* **7**, e49958. (doi:10.1371/journal.pone.0049958)
35. Boyle M, DiNardo S. 1995 Specification, migration and assembly of the somatic cells of the *Drosophila* gonad. *Development* **121**, 1815–1825.
36. King RC. 1970 *Ovarian development in Drosophila melanogaster*, p. 227. New York, NY: Academic Press.
37. Sahut-Barnola I, Godt D, Laski FA, Couderc J-L. 1995 *Drosophila* ovary morphogenesis: analysis of terminal filament formation and identification of a gene required for this process. *Dev. Biol.* **170**, 127–135.
38. Hodin J, Riddiford LM. 1998 The ecdysone receptor and *ultraspiracle* regulate the timing and progression of ovarian morphogenesis during *Drosophila* metamorphosis. *Dev. Genes Evol.* **208**, 304–317. (doi:10.1007/s004270050186)
39. Gancz D, Lengil T, Gilboa L. 2011 Coordinated regulation of niche and stem cell precursors by hormonal signaling. *PLoS Biol.* **9**, e1001202. (doi:10.1371/journal.pbio.1001202)
40. Fernandez R, Tabarini D, Azpiazu N, Frasch M, Schlessinger J. 1995 The *Drosophila* insulin receptor homolog: a gene essential for embryonic development encodes two receptor isoforms with different signaling potential. *EMBO J.* **14**, 3373–3384.
41. Brogiolo W, Stocker H, Ikeya T, Rintelen F, Fernandez R, Hafen E. 2001 An evolutionarily conserved function of the *Drosophila* insulin receptor and insulin-like peptides in growth control. *Curr. Biol.* **11**, 213–221. (doi:10.1016/S0960-9822(01)00068-9)
42. Chen C, Jack J, Garofalo RS. 1996 The *Drosophila* insulin receptor is required for normal growth. *Endocrinology* **137**, 846–856.
43. Wu Q, Zhang Y, Xu J, Shen P. 2005 Regulation of hunger-driven behaviors by neural ribosomal S6 kinase in *Drosophila*. *Proc. Natl Acad. Sci. USA* **102**, 13 289–13 294. (doi:10.1073/pnas.0501914102)
44. Manseau L *et al.* 1997 GAL4 enhancer traps expressed in the embryo, larval brain, imaginal discs, and ovary of *Drosophila*. *Dev. Dyn.* **209**, 310–322. (doi:10.1002/(SICI)1097-0177(199707)209:3<310::AID-AJA6>3.0.CO;2-L)
45. Catchpole RDJ. 1994 Wing length is not the best predictor of body size. *Drosoph. Inf. Serv.* **75**, 84–86.
46. Puig O, Marr MT, Ruhf ML, Tjian R. 2003 Control of cell number by *Drosophila* FOXO: downstream and feedback regulation of the insulin receptor pathway. *Genes Dev.* **17**, 2006–2020. (doi:10.1101/gad.1098703)
47. Hietakangas V, Cohen SM. 2009 Regulation of tissue growth through nutrient sensing. *Annu. Rev. Genet.* **43**, 389–410. (doi:10.1146/annurev-genet-102108-134815)
48. Tang HY, Smith-Caldas MSB, Driscoll MV, Salhadar S, Shingleton AW. 2011 FOXO regulates organ-specific phenotypic plasticity in *Drosophila*. *PLoS Genet.* **7**, e1002373. (doi:10.1371/journal.pgen.1002373)
49. Emlen DJ, Warren IA, Johns A, Dworkin I, Lavine LC. 2012 A mechanism of extreme growth and reliable signaling in sexually selected ornaments and weapons. *Science* **337**, 860–864. (doi:10.1126/science.1224286)
50. Snell-Rood EC, Moczek AP. 2012 Insulin signaling as a mechanism underlying developmental plasticity: the role of FOXO in a nutritional polyphenism. *PLoS ONE* **7**, e34857. (doi:10.1371/journal.pone.0034857)
51. Yano H, Nakanishi S, Kimura K, Hanai N, Saitoh Y, Fukui Y, Nonomura Y, Matsuda Y. 1993 Inhibition of histamine secretion by wortmannin through the blockade of phosphatidylinositol 3-kinase in RBL-2H3 cells. *J. Biol. Chem.* **268**, 25 846–25 856.
52. Legrand D, Tenailon MI, Matyot P, Gerlach J, Lachaise D, Cariou M-L. 2009 Species-wide genetic variation and demographic history of *Drosophila sechellia*, a species lacking population structure. *Genetics* **182**, 1197–1206. (doi:10.1534/genetics.108.092080)
53. R'Kha S, Capy P, David JR. 1991 Host-plant specialization in the *Drosophila melanogaster* species complex: a physiological, behavioral, and genetic analysis. *Proc. Natl Acad. Sci. USA* **88**, 1835–1839. (doi:10.1073/pnas.88.5.1835)
54. Lachaise D, Cariou M-L, David JR, Lemeunier F, Tsacas L, Ashburner M. 1988 Historical biogeography of the *Drosophila melanogaster* species subgroup. *Evol. Ecol.* **22**, 159–225.
55. Engstrom LE. 1971 *Studies of the effects of two-way selection for ovariole number in Drosophila melanogaster*. Urbana-Champaign, IL: University of Illinois at Urbana-Champaign.
56. Tatar M, Kopelman A, Epstein D, Tu MP, Yin CM, Garofalo RS. 2001 A mutant *Drosophila* insulin receptor homolog that extends life-span and impairs neuroendocrine function. *Science* **292**, 107–110. (doi:10.1126/science.1057987)
57. Clancy DJ, Gems D, Harshman LG, Oldham S, Stocker H, Hafen E, Leevers SJ, Partridge L. 2001 Extension of life-span by loss of CHICO, a *Drosophila* insulin receptor substrate protein. *Science* **292**, 104–106. (doi:10.1126/science.1057991)
58. Paaby AB, Blacket MJ, Hoffmann AA, Schmidt PS. 2010 Identification of a candidate adaptive polymorphism for *Drosophila* life history by parallel independent clines on two continents. *Mol. Ecol.* **19**, 760–774. (doi:10.1111/j.1365-294X.2009.04508.x)
59. Sutter NB *et al.* 2007 A single IGF1 allele is a major determinant of small size in dogs. *Science* **316**, 112–115. (doi:10.1126/science.1137045)